

## Determination of Cyclopiazonic Acid in Chinese Yellow Wine by High-Performance Liquid Chromatography–Triple Quadrupole Mass Spectrometry

<sup>1</sup>Qi Peng, <sup>1</sup>Wenhao Xing, <sup>1</sup>Qinxia Xu, <sup>1</sup>Jiaying Chen, <sup>1</sup>Hao Yu, <sup>2</sup>Feiran Chen, <sup>2</sup>Bobin Li, <sup>1</sup>Xia Xu, <sup>1</sup>Zhongyuan Wang, <sup>1</sup>Rungang Tian, <sup>1</sup>Jianqiu Sun, <sup>3</sup>HuiJun Zou, <sup>1</sup>Yiwei Mo\* and <sup>3</sup>Guangfa Xie\*

<sup>1</sup>School of Life Sciences, Shaoxing University, Shaoxing, 312000 China.

<sup>2</sup>Shaoxing Testing Institute of Quality Technical Supervision, Shaoxing, 312071 China.

<sup>3</sup>National Engineering Research Center for Chinese Rice Wine, China Shaoxing Rice Wine Group Co., Ltd, Shaoxing 312000, China.

mike.peng@126.com\*

(Received on 29<sup>th</sup> June 2016, accepted in revised form 2<sup>nd</sup> February 2017)

**Summary:** We developed a detection method of cyclopiazonic acid (CPA), a mycotoxin in Chinese yellow rice wine, based on high-performance liquid chromatography and triple quadrupole mass spectrometry. Under optimized conditions, the limit of detection was 50 µg/L. CPA recovery rates in CPA-spiked Chinese yellow rice wine were 80.1% and 85.1% at 25 µg/L and 1,050 µg/L CPA, respectively ( $R > 0.9994$ ). The relative standard deviation was 6.21–9.17%. Our developed method represents a rapid and accurate detection tool of CPA meeting minimum residue measurement requirements in Chinese yellow rice wine. The method will be widely available as a reference and be employed for Chinese official regulatory control purposes.

Keywords: Cyclopiazonic acid; HPLC/QQQ MS; Rice Wine

### Introduction

Cyclopiazonic acid (CPA), an indole tetramic acid, is a toxic fungal secondary metabolite [1,2] produced by *Penicillium cyclopium*, *P. griseofulvum*, *P. camemberti*, *P. commune*, *Aspergillus flavus*, and *A. versicolor*. Studies have reported that this mycotoxin is present in grains, dried beans, grain-based products, and livestock feed contaminated with *Penicillin* and *Aspergillus* strains. While CPA and aflatoxin (total aflatoxins) are often detected in similar environmental conditions, CPA is more toxic [3-6]. According to animal studies, the oral administration of CPA to rats causes degenerative changes and necrosis to multiple organs (liver, spleen, exocrine and endocrine pancreas, kidney, salivary glands, myocardium, and skeletal muscle) and death in 1 – 5 d [7]. In rats, CPA has an LD<sub>50</sub> of 2.3 mg/kg [8].

Chinese yellow rice wine is a traditional fermented wine, which is consumed as a beverage and used in food seasoning. Inappropriate practices during yellow rice wine fermentation processes increase the risk of fungal growth and toxin formation. Currently, common CPA detection methods include immunoassays [4], capillary electrophoresis [9], reverse ligand exchange chromatography [10], silica gel column chromatography [11], ion chromatography [12], and metal complex chromatography [13, 14]. However, the extraction steps of these methods are complicated, time-consuming, expensive, and require large volumes of toxic reagents such as chloroform or dichloromethane [15, 16].

In this study, we used high performance

liquid chromatography (HPLC) as a CPA separation method and triple quadrupole mass spectrometry (QQQ MS) as a CPA detection method. Under optimized conditions, we measured the level of the mycotoxin in Chinese yellow rice wine.

### Experimental

#### Materials and Instrumentation

All chemicals used in this study were of HPLC grade (CNW Technologies GmbH, Germany). CPA (purity >97%) was purchased from Sigma-Aldrich. Chinese yellow rice wine was obtained from a local market in Shaoxing city.

HPLC was performed with an Ultimate C18-WP column (5 µm, 4.6 mm × 250 mm, CNW) maintained at 35°C. The injection volume was 10 µl, and the mobile phases included A (aqueous 0.05 mol/L ammonium acetate: glacial acetic acid adjusted to pH 6.50 ± 0.05) and B (100% methanol). The A-to-B mobile phase ratio was 30/70 (v/v), with a flow rate of 0.3 ml/min.

Mass spectrometry was performed in an Agilent 1290 LC/MSD TraCp 6460 (CA, USA). QQQ MS conditions included an electrospray ionization (ESI) in negative mode, an atomizing air pressure of 275.8 kPa, a drying gas (N<sub>2</sub>) flow rate of 11 L/min at 350°C, a nebulizer pressure of 50 PSI, a capillary voltage of 3,500 V, a sheath gas temperature of 350°C, and a nozzle voltage of 500 V. Selected reaction monitoring was used, with quantitative and qualitative ion pairs at m/z 335, 154 and 335, 140, respectively. Centrifugation was performed in a Hermle Model Z. 200 A (Wehingen, Germany).

\*To whom all correspondence should be addressed.

Filtered, deionized water was obtained from a Milli-Q ultrapure water system (America Millipore). Other instruments included a MTN-2800D nitrogen concentrator (Tianjin Auto Science, China) and a HC-C18 solid phase extraction column (CNW, Germany).

#### Sample preparation

Chinese yellow rice wine (5 ml) was homogenized for 120 min with 5 ml of extraction solution (70% methanol and 30% of 1% NaHCO<sub>3</sub>) and centrifuged for 15 min at 1,000×g. An aliquot (4 ml) of the resulting supernatant was mixed with 10 ml of hexane to remove lipids and centrifuged for 5 min at 10,000×g. This supernatant was diluted further with 20mL water and loaded on a solid phase extraction cartridge that had been prepared for use. Vacuum was employed to draw the sample through the cartridge at a flow rate of approximately 1mL per minute. The cartridge was dried with a stream of nitrogen, and the analytes were eluted with 5mL water/ methanol (1:1, v/v) at 1mL/min. An anion exchange column, a strong cation exchange column, and a reverse phase column were evaluated respectively. The solvent was evaporated under a stream of nitrogen gas at 50°C [17]. The internal standard was added in 1mL of mobile phase and the residue was dissolved with vortex mixing. The dissolved extract was passed through a microporous filter membrane (0.22 μm) (Millipore Corp., Bedford, Mass.) for HPLC/QQQ MS analysis. [17, 18]

## Results and Discussion

#### Optimization of CPA purification

Immunoaffinity columns are ideal for the purification of specific CPAs. However, there are no suitable immunoaffinity columns for CPA purification. Therefore, we evaluated the performance (i.e., retention and recovery) of different columns, including an anion exchange column, a strong cation exchange column, and a reverse phase column. The results revealed that CPA recovery is 85% (77%), 73% (70%) and 69% (63%) respectively by anion exchange column, cation exchange column, and reverse phase column (Table-1). The anion exchange column was the most optimum column for CPA purification. The CPA phenolic groups formed strong interactions with the quaternary ammonium groups on the anion exchange column.

#### Optimization of HPLC/QQQ MS conditions

Two mobile phases were investigated in this study: aqueous acetonitrile and aqueous methanol. The highest analytical signal was obtained with aqueous ammonium acetate/methanol as the mobile phase. The addition of 0.005% aqueous ammonium

acetate to the aqueous methanol phase increased signal strength. Matrix of Chinese yellow rice wine effects can be predicted. It is highly variable and difficult to control. The suppression or enhancement may be severity and nature for function of the concentration of the co-eluting matrix components. [10, 19].

Table-1: Optimization of CPA recovery.

Name	Recovery (%)	
	Elution with MeOH	Elution with Acetonitrile
Anion exchange column	85	77
Cation exchange column	73	70
Reverse phase column	69	63

Precursor and product ions of CPA were selected by full scan mass spectrometry using positive and negative electrospray ionization. Improved sensitivity was obtained in the negative ion mode. QQQ MS analyzes identified two characteristic peaks (m/z 154 and m/z 140). High abundance ions were selected as quantitative (m/z 335 >154) and qualitative (m/z 335 >140) ions. Selective ion monitoring mass spectrometry provided a signal-to-noise area ratio of 3 and ensured accuracy in the results.

#### Validation of the HPLC/QQQ MS method

The calibration curve (Fig. 1) generated with different CPA standard concentrations (25, 50, 250, 350, and 1,050 μg/L) had a regression equation of  $Y = 12.717886X - 184.530097$  ( $R^2 = 0.9994$ ). Limit of detection (LOD) and limit of quantification (LOQ) were measured based on a signal-to-noise ratio of 3 and 10, respectively. In this study, LOD was 50 μg/L and LOQ was 240 μg/L. To determine recovery rates, four blanks samples were each spiked with 50, 100, 200, and 500 μg/L CPA. The recovery rates were 80.1–85.1% (Table-2), with a relative standard deviation of 6.21% – 9.17%.

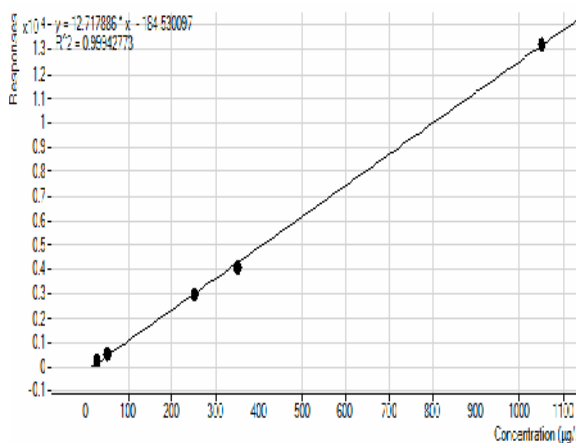


Fig. 1: Calibration curve for the cyclopiazonic acid.

Table-2: Recovery rates of cyclopiazonic acid (CPA) from CPA-spiked Chinese yellow rice wine.

Sample	Aging time (months)	Concentration of CPA added (µg/L)	Average recovery rate (%)	RSD (%)
1	12	50	80.1	6.21
	24	50		
	36	50		
2	12	100	83.2	7.77
	24	100		
	36	100		
3	12	200	85.1	8.19
	24	200		
	36	200		
4	12	200	81.9	9.17
	24	200		
	36	200		

RSD: relative standard deviation. Recovery experiments (n= 6)

Analysis of Chinese Yellow Rice Wine

Yellow rice wine purchased at a local Chinese market was analyzed using our developed method. The results revealed that the commercial yellow rice wine was CPA-negative. Yellow rice wine was spiked with 200 µg/L CPA and analyzed by HPLC/QQQ MS. The results showed that a visible quantitative ion peak, with a qualitative ion-to-quantitative ion relative abundance of <20% (Fig. 2). It is suitable for its intended test purpose. The goal of validation of the analytical procedure was demonstrated by analysis of spiked Chinese yellow rice wine. [19]

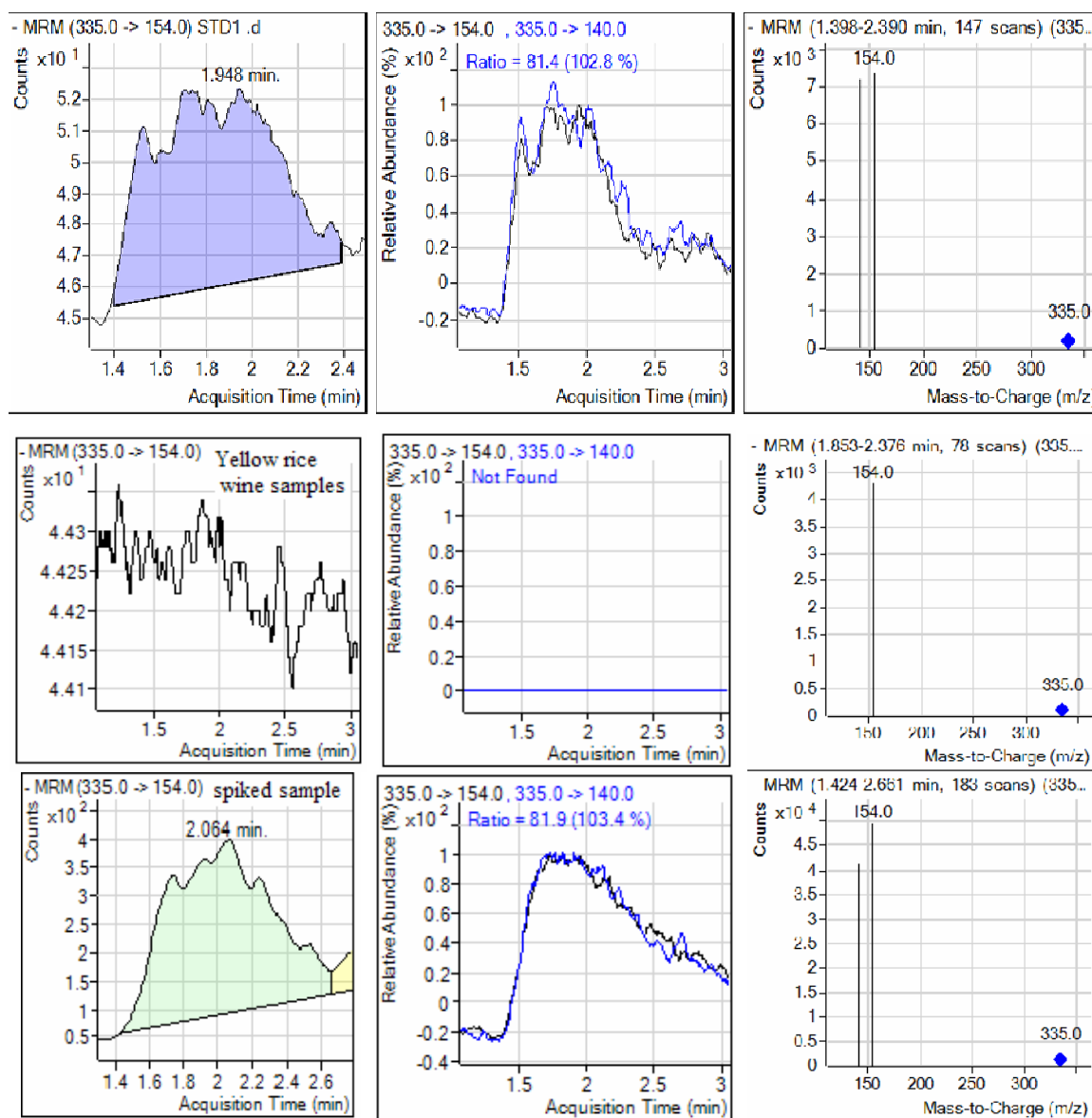


Fig. 2: Triple quadrupole mass spectrometry (QQQ MS) of CPA in commercial Chinese yellow rice wine.

## Conclusions

Our validated method has sufficient sensitivity to detect the presence of CPA in Chinese yellow rice wine. Even though no maximum residue limits have been established, our method met the required performance criteria set by the Chinese Food and Drug Administration. The precision was good and the recovery rates were within the permissible range (80.1 – 85.1%). Therefore, our developed method can be used in the analyses of yellow rice wine and to support food safety agencies for regulatory control purposes.

## Acknowledgements

We thank China Food and Drug Administration for providing samples and the financial support provided by Foundation of Public Projects of Zhejiang Province, China (No. 2017C32101).

## References

1. N. Agata, H. Tanaka and K. Shigenobu, Possible Action of Cyclopiazonic Acid on Myocardial Sarcoplasmic Reticulum: Inotropic Effects on Neonatal and Adult Rat Heart, *J. Br J Pharmacol.*, **10**, 571 (1993).
2. A. Antonella, C. Nicola and P. Francesco, Simultaneous Determination of Ochratoxin A and Cyclopiazonic, Mycophenolic, and Tenuazonic Acids in Cornflakes by Solid-Phase Microextraction Coupled to High- Performance Liquid Chromatography, *J. Agric. Food Chem.*, **51**, 5232 (2003).
3. R. B. Lalitha and A. Husain, Presence of Cyclopiazonic Acid in Kodo Millet (*Paspalum scrobiculatum*) causing Kodua Poisoning in Man and its Production by Associated Fungi, *J. Mycopathologia.*, **89**, 177 (1985).
4. L. R. Monaci, G. E. Vatinno, and D. E. Benedetto, Fast Detection of Cyclopiazonic Acid in Cheese Using Fourier Transform Mid-Infrared ATR Spectroscopy, *J. Eur. Food Res. Technol.*, **225**, 585 (2007).
5. M. Sánchez-Hervás, J. V. Gil, F. Bisbal, D. Ramón and P. V. Martínez-Culebras, Mycobiota and Mycotoxin Producing Fungi from Cocoa Beans, *J. Food Microbiol Int.*, **125**, 336 (2008).
6. M. Takemoto, K. Takagi, and K. Ogino, Comparison of Contractions Produced by Carbachol, Thapsigargin and Cyclopiazonic Acid in the Guinea-Pig Tracheal Muscle, *J. Br J Pharmacol.*, **124**, 1449 (1998).
7. I. F. H. Purchase, The Acute Toxicity of the Mycotoxin Cyclopiazonic Acid to Rats, *Toxicol. Appl. Pharmacol.*, **18**, 114 (1971).
8. J. E. Hill, L. G. Lomax and R. J. Gole, Toxicologic and Immunologic Effects of Sublethal Doses of Cyclopiazonic Acid in Rat, *J. Am J Vet Res.*, **47**, 1174 (1986).
9. Y. Hayashi, T. Yoshizawa, Analysis of Cyclopiazonic Acid in Corn and Rice by a Newly Developed Method, *J. Food Chem.*, **93**, 215 (2005).
10. R. Widiastuti, R. Maryam, and B. J. Blaney, Cyclopiazonic Acid in Combination with Aflatoxins, Zearalenone and Ochratoxin A in Indonesian Corn, *J. Mycopathologia.*, **104**, 153 (1988).
11. J. D. Bailly, C. Tabuc and A. Querin, Production and Stability of Pa-Tulin, Ochratoxin A, Citrinin, and Cyclopiazonic Acid on Dry-Cured Ham, *J. Food Prot.*, **68**, 1516 (2005).
12. M. J. Sosa, J. J. Cordoba and C. Diaz, Production of Cyclopiazonic Acid by *Penicillium Commune* Isolated from Dry-Cured Ham on A meat Extract-Based Substrate, *J. Food Prot.*, **65**, 988 (2002).
13. N. Gqaleni, E. Smith and J. Lacey, Effects of Temperature, Water Activity and Incubation Time on Production of Aflatoxins and Cyclopiazonic Acid by an Isolate of *Aspergillus Flavus* in Surface Agar Culture, *J. App. Environ. Microbiol.*, **63**, 1048 (1997).
14. A. S. Moldes-Anaya, T. N. Asp, and G. S. Eriksen, Determination of Cyclopiazonic Acid in Food and Feeds by Liquid Chromatography-Tandem Mass Spectrometry, *J. Chromatogr A.*, **1216**, 3812 (2009).
15. V. S. Sobolev, Determination of Ionization Constant (pKa) and Octanol-Water Partition Coefficient (Log P) of Cyclopiazonic Acid, *J. A. O. A. C. Int.*, **88**, 1367 (2005).
16. M. L. Martins and H. M. Martins, Natural and Invitro Coproduction of Cyclopiazonic Acid and Aflatoxins, *J. Food Prot.*, **62**, 292 (1999).
17. Q. Peng, R. G. Tian, B. B. Li, J. G. Hu, Y. Q. Meng and J. L. Wang, Determination of Luteoskyrin in Rice Wine by High-Performance Liquid Chromatography-Ion Trap Tandem Mass Spectrometry, *J. Anal Lett.*, **48**, 88 (2015).
18. C. W. Holzapfel, The Isolation and Structure of Cyclopiazonic Acid, a Toxic Metabolite of *Penicillium Cyclopium* Westling, *J. Tetrahedron.*, **24**, 2101 (1968).
19. R. Czernych, J. J. Halkiewicz, A. Kot-Wasik, and J. Namieśnik, Development and Validation of SPE-HPLC-MS/MS Method for Determining Cyclo- Phosphamide in Surface Waters. Pol, *J. Environ. Stud.*, **5**, 23 (2014)